

AMPHETAMINES IN BLOOD, PLASMA/SERUM, URINE, OR TISSUE USING CLEAN SCREEN® DAU SPE AND LC-MS/MS ANALYSIS

### **UCT Part Numbers**

CSDAU206 Clean Screen® DAU 200 mg, 6 mL Column

**SLPFPP50ID2.1-1.8UM** Selectra<sup>®</sup> PFPP UHPLC Column 50 X 2.1 mm, 1.8 μm

**SLPFPPGDC20-1.8UM** Selectra® PFPP Guard Column 10 X 2.0 mm, 1.8 μm

> **SLGRDHLDR-HP** Guard Column Holder



FORENSICS



## **Summary:**

Amphetamines are a group of drugs that stimulate the central nervous system (CNS). These drugs can be administered in the body through many ways. While oral consumption is the most common route, they also can also be snorted, smoked and injected intravenously. Increasing abuse potential and dependence liability of amphetamine & methamphetamine have caused the DEA/FDA to classify these drugs as Schedule II controlled substances. The ease of manufacturing has made methamphetamine one of the most frequently encountered substance in related cases. drug Designer drugs metylenedioxymethamphetamine (MDMA) and methylenedioxyamphetamine methylenedioxy derivatives of methamphetamine (MDA) are and amphetamine respectively. Phentermine is a schedule IV drug that is not heavily abused but is known to exert effects that are similar to Amphetamine.

This application note describes a simple and robust solid-phase extraction (SPE) procedure for amphetamines in blood, plasma/serum, urine and tissue samples. The mixed-mode functionality of the Clean Screen® DAU SPE cartridge ensures efficient extraction of the amphetamines while removing undesired matrix components and yielding clean extracts. UHPLC separation was carried out using a Selectra® PFPP column prior to detection by tandem mass spectrometry (MS/MS). The PFPP (pentafluorophenylpropyl) stationary phase can undergo dipole-dipole and pi-pi interactions, imparting unique selectivity and retention mechanisms to the column that distinguish it from other stationary phases. In this application excellent retention of the polar amphetamines, including baseline separation of the isobaric methamphetamine and phentermine, was obtained in less than 4.5 minutes.

## **SPE Procedure:**

#### 1) Sample Preparation:

- Add appropriate volumes of internal standard to 1 -2 mL of blood, plasma/ serum, urine, or 1 g (1:4) tissue homogenate
- Mix/vortex briefly and let stand for 5 minutes
- Add 3 mL of 100 mM phosphate buffer (pH 6.0)
- Mix/vortex briefly
- For blood, plasma/ serum tissue homogenate samples, centrifuge for 10 minutes at 2000 rpm (discard pellet after loading sample onto SPE cartridge)

#### 2) Condition SPE Sorbent:

- 1 x 3 mL methanol
- 1 x 3 mL 100 mM phosphate buffer (pH 6.0)

#### 3) Apply sample:

• Load sample at 1-2 mL/minute

#### 4) Wash Sorbent:

- 1 x 3 mL 0.1 M HCl
- 1 x 3 mL methanol

#### 5) Dry Sorbent:

• Dry SPE cartridge for 2 mins at 80-100 psi

#### 6) Elute:

- 1 x 3 mL ethyl acetate/ IPA/ NH<sub>4</sub>OH (78:20:2)
- Collect eluate at 1-2 mL/minute

#### 7) Evaporate Eluent:

- Evaporate the eluent for 5 minutes to remove  $NH_4OH$  (40°C, gentle stream of  $N_2$ )
- Add 100  $\mu\text{L}$  of 1% HCl in methanol to prevent volatization of the drugs and loss during evaporation

**Note:** it is important to remove the NH<sub>4</sub>OH prior to adding 1% HCl in methanol, otherwise a white precipitate (NH<sub>4</sub>Cl) will form.

#### 8) Reconstitute:

• Reconstitute samples in 100 μL of mobile phase (alternative volumes may also be used)





# LC-MS/MS Parameters:

System: Shimadzu LCMS-8050						
UHPLC Column: Selectra <sup>®</sup> PFPP (50 X 2.1 mm, 1.8 μm)						
Guard Column: Selectra <sup>®</sup> PFPP (10 X 2.0 mm, 1.8 μm)						
Column Temperature: 40°C						
Flow Rate: 0.5 mL/min						
Injection Volume: 2 μL						
Autosampler temperature: 10°C						
Gradient Program:						
Time (min)	% Mobile Phase A (0.1% Formic Acid in Water)	% Mobile Phase B (0.1% Formic Acid in Methanol)				
<b>Time (min)</b> 0.0						
	(0.1% Formic Acid in Water)	(0.1% Formic Acid in Methanol)				
0.0	(0.1% Formic Acid in Water) 100	(0.1% Formic Acid in Methanol)				
0.0	(0.1% Formic Acid in Water) 100 70	(0.1% Formic Acid in Methanol) 0 30				
0.0 0.5 3.0	(0.1% Formic Acid in Water) 100 70 60	(0.1% Formic Acid in Methanol) 0 30 40				
0.0 0.5 3.0 3.5	(0.1% Formic Acid in Water) 100 70 60 0	(0.1% Formic Acid in Methanol) 0 30 40 100				

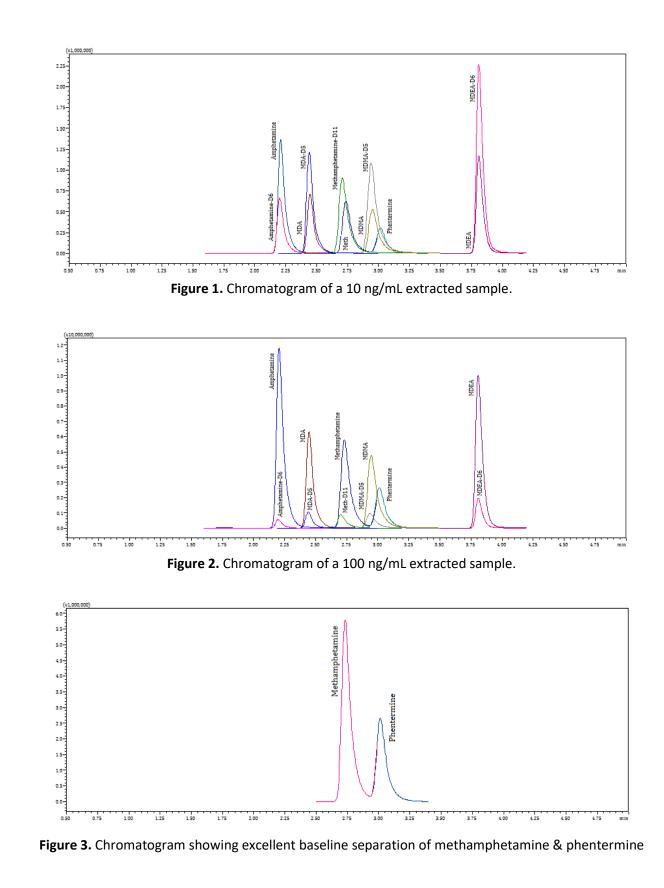
## **Results:**

Recovery - Blood					
Analyte	10 ng/mL (n=3)	Rel. Std Dev (%)	100 ng/mL (n=3)	Rel. Std Dev (%)	
Amphetamine	96%	2.55	92%	0.78	
Methamphetamine	95%	2.59	92%	1.72	
Phentermine	104%	3.37	95%	8.16	
MDA	100%	4.80	94%	1.16	
MDMA	97%	3.36	94%	0.64	
MDEA	93%	1.87	91%	0.36	
Recovery - Urine					
Analyte	10 ng/mL (n=3)	Rel. Std Dev (%)	100 ng/mL (n=3)	Rel. Std Dev (%)	
Amphetamine	104%	2.85	95%	2.02	
Methamphetamine	103%	3.38	93%	3.03	
Phentermine	117%	4.08	107%	5.25	
MDA	106%	2.27	96%	3.04	
MDMA	105%	2.82	96%	2.62	
MDEA	102%	2.16	94%	2.41	





## **Chromatograms:**







### **Calibration Curves:**

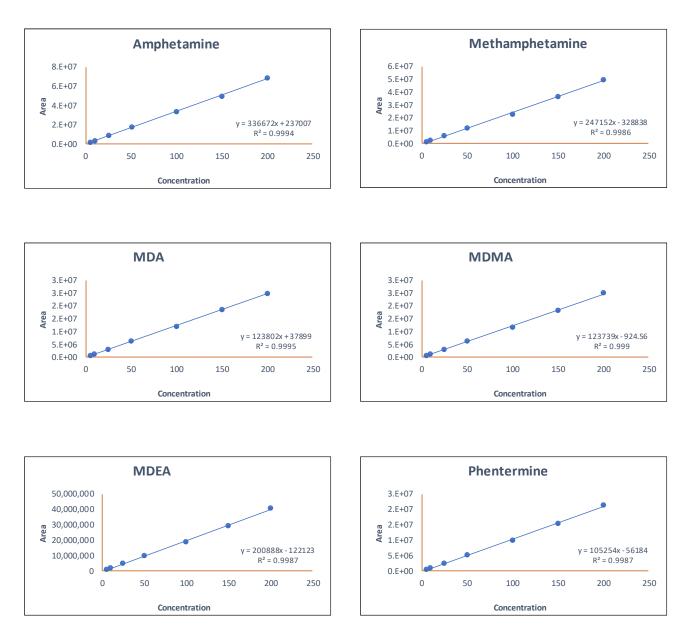


Figure 4. Calibration curve for the six amphetamines (5, 10, 25, 50, 100, 150 & 200 ng/mL).

**Note:** For accurate quantitation of recoveries and to prevent saturation of the MS detector, a calibration curve ranging from 5-200 ng/ml was utilized for this study. Depending upon the requirements of an individual testing lab, a calibration curve with a wider concentration range may be required for routine analysis.





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